



An illustrated key to the genera and subgenera of the Alysiini (Hymenoptera, Braconidae, Alysiinae), with three genera new for China

Jia-Chen Zhu¹, Cornelis van Achterberg^{2,3}, Xue-Xin Chen¹

I State Key Laboratory of Rice Biology and Ministry of Agriculture Key Lab of Agricultural Entomology, Institute of Insect Sciences, Zhejiang University, Hangzhou 310058, China 2 Key Laboratory of Resource Biology and Biotechnology in Western China (Northwest University) 3 Ministry of Education, College of Life Sciences, Northwest University, 229 North Taibai Road, Xi'an, Shaanxi 710069, China

Corresponding author: Xue-xin Chen (xxchen@zju.edu.cn)

Academic editor: M. Sharkey | Received 20 June 2017 | Accepted 6 November 2017 | Published 13 December 2017

http://zoobank.org/59E9E632-01B1-4503-BCAC-1465F9ADC436

Citation: Zhu J-C, van Achterberg C, Chen X-X (2017) An illustrated key to the genera and subgenera of the Alysiini (Hymenoptera, Braconidae, Alysiinae), with three genera new for China. ZooKeys 722: 37–79. https://doi.org/10.3897/zookeys.722.14799

Abstract

An illustrated key to the genera and subgenera of the Alysiini (Hymenoptera, Braconidae, Alysiinae) from China is presented. Three genera new for China are reported: *Adelurola* Strand, 1924, *Anisocyrta* Foerster, 1863, and *Pentapleura* Foerster, 1863. The total for China is 26 genera of Alysiini and an additional seven subgenera (excluding the nominal subgenera, which are included in the total of genera). The known Chinese species are listed under each genus and the biology is summarised. *Separatatus sinicus* (Zheng, Chen & Yang, 2012) and *Grammospila eurys* (Chen & Wu, 1994) are new combinations. *Regetus* Papp, 1999, and *Adelphenaldis* Fischer, 2003, are new synonyms of *Eusynaldis* Zaykov & Fischer, 1982. In addition, *Eusynaldis* Zaykov & Fischer and *Synaldis* Foerster, 1863, are treated as subgenera of *Aspilota* Foerster, 1863, and *Dinotrema* Foerster, 1863, respectively. An aberrant species of *Separatatus* Chen & Wu, 1994, *S. parallelus* **sp. n.**, is described from Yunnan and Hainan.

Keywords

Alysiinae, Alysiini, Braconidae, China, Hymenoptera, key to genera, new record, Oriental, Palaearctic

Introduction

The subfamily Alysiinae Leach, 1815 (Hymenoptera: Braconidae) contains small to medium-sized koinobiont endoparasitoids of cyclorrhaphous dipterous larvae (Wharton 1984; Shaw and Huddleston 1991; van Achterberg 1993). Alysiinae is characterized among the Braconidae by having exodont mandibles, a feature occurring almost exclusively in this subfamily. The mandibles do not touch each other, even when they are closed (van Achterberg 1993; Belokobylskij and Kostromina 2011). Specimens of Alysiinae are often common, especially when decaying organic material is abundant (Peris-Filipo and Jimenez-Peydro 2011; pers. obs.).

Keys to the genera of Alysiinae of the Old World are found in Fischer (1976a) (including all known genera up to 1975), Chen and Wu (1994) (key to genera of China) and Wharton (2002) (key to genera of the Australian region). All of these keys are useful, but are not illustrated and do not include all the genera found during our study. Therefore, an illustrated key to all genera and subgenera of the Alysiini known from China is presented in this paper.

Chen and Wu (1994) reported 19 genera and *Heterolexis* Foerster as a subgenus, but the report of *Adelurola* Strand is not accepted because the included species belongs to *Grammospila* Foerster. Wu et al. (1995a) and Yao (2015b) reported *Cratospila* Foerster, and *Trachyusa* Ruthe, respectively. Zheng et al. (2012) added *Bobekoides* van Achterberg, but the reported species is here transferred to *Separatatus* Chen & Wu. Chen and Wu (1994) indirectly reported *Grammospila* (because of the reported species) and the subgenera *Eusynaldis* Zaykov & Fischer and *Synaldis* Foerster. These subgenera are recognised for convenience, because their recognition likely renders the genera *Aspilota* Foerster and *Dinotrema* Foerster paraphyletic. Recently, the total number of genera for China reached 23 by the publication of *Dacnulysia* Zhu, van Achterberg & Chen by Zhu et al. (2017).

In this paper three genera are listed as new for China: *Adelurola* Strand, *Anisocyrta* Foerster and *Pentapleura* Foerster. The total for China is 26 genera of Alysiini and seven subgenera (without the nominal subgenera; they are included in the total of genera), comprising 132 species.

Materials and methods

The collection specimens were hand net collected and glued on card points. They were sorted from the Braconidae collection present in the Institute of Insect Sciences of the Zhejiang University (**ZJUH**). The terminology and measurements used follow van Achterberg (1979, 1988a). The following abbreviations are used: **POL** – postocellar line; **OOL** – ocular-ocellar line, measured from ocellus directly to eye; **OD** – maximum diameter of lateral ocellus; medial length of the first tergite is measured from the apex of the adductor to the apex of tergite. Descriptions and measurements were made under a Leica M125 stereomicroscope. Photographs were made with a Keyence

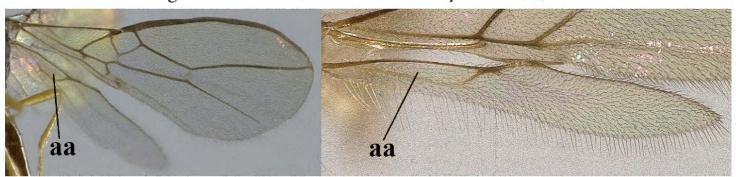
VHX-2000 digital microscope and the photos were slightly processed (mainly cropped and backgrounds modified) in Photoshop CC. The drawings are from van Achterberg (1988b). The literature on Chinese Alysiini and the original publications of the genera are referenced; for additional references, see Yu et al. (2016).

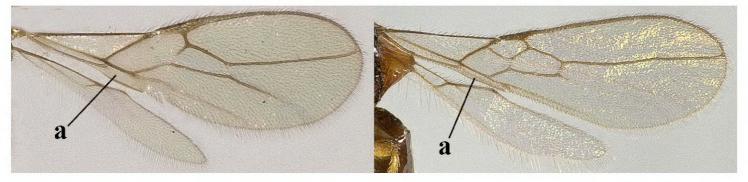
Key to genera of Chinese Alysiini

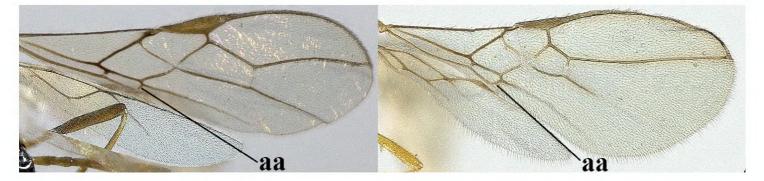
1 Hind wing without closed cells and very narrow (a); [few aberrant spp.]......

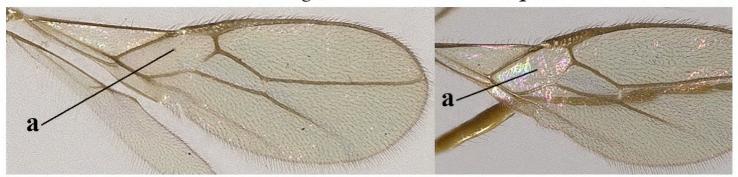
Dinotrema Foerster, 1863 p.p.









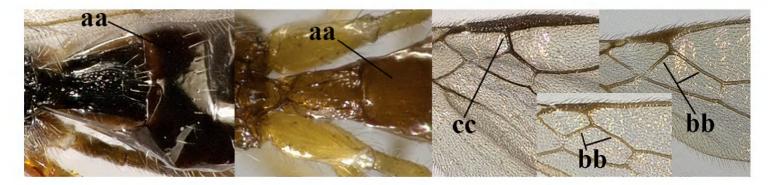


Vein 1-SR+M of fore wing present (aa)4

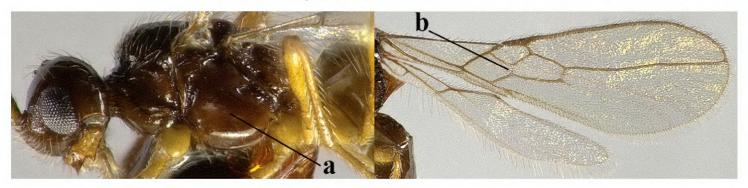




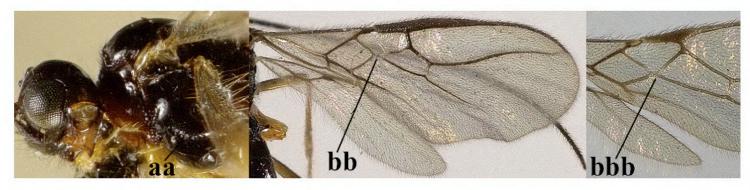
Second tergite smooth (aa); vein 2-SR of fore wing shorter than vein 3-SR
 (bb) or vein r of fore wing emitted near basal third of pterostigma (cc)......5

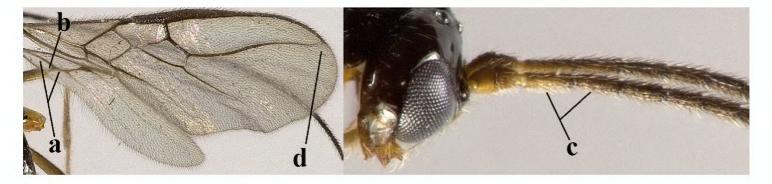


Precoxal sulcus absent (a), at most shallowly impressed and with some micro-sculpture; vein m-cu of fore wing (just) postfurcal (b) Pentapleura Foerster, 1863

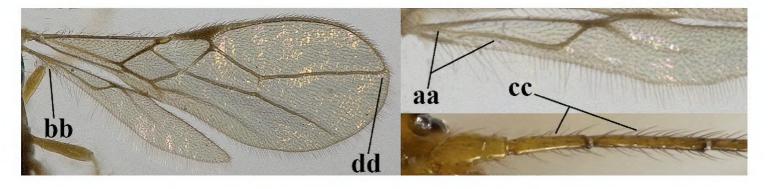


Precoxal sulcus at least medially distinctly impressed and with some (micro-) crenulae (aa); vein m-cu of fore wing antefurcal (bb) or interstitial (bbb)....6

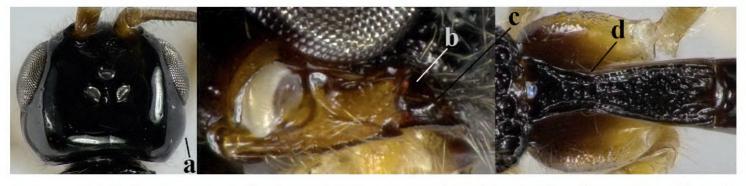




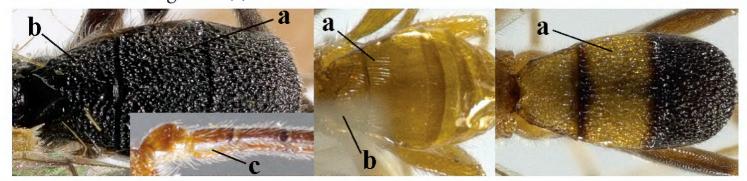
Vein M+CU of hind wing distinctly shorter than vein 1-M (aa) or vein cu-a absent (bb); third antennal segment usually shorter than fourth segment (cc); marginal cell of fore wing reaching wing apex (dd)..... Asobara Foerster, 1863

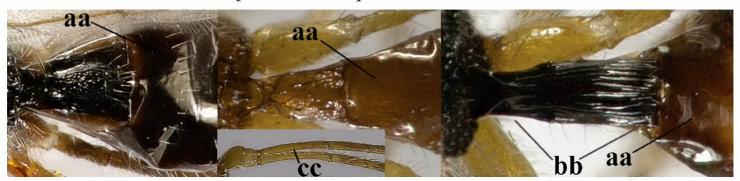


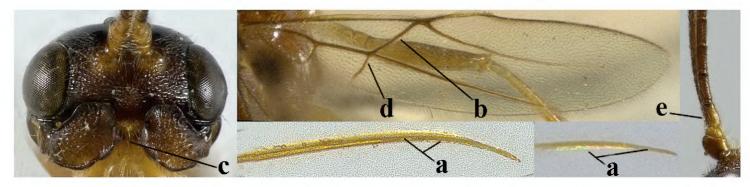
Head nearly square in dorsal view (a); mandible with wide gap between first and second tooth (b) and second tooth with dorsal tooth (c); first metasomal tergite (compared to base of tergite) distinctly constricted near basal third (d); [metasoma of \$\to\$ compressed; first tergite without dorsope, except elongate shallow depression (d)] *Dacnulysia* Zhu, van Achterberg & Chen, 2017

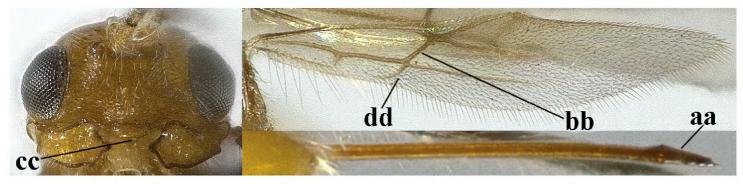


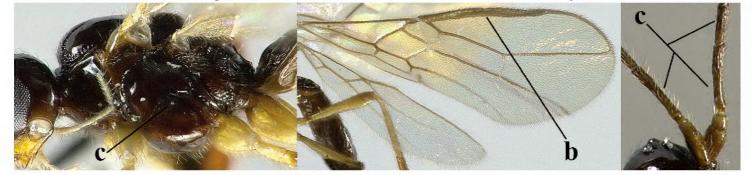




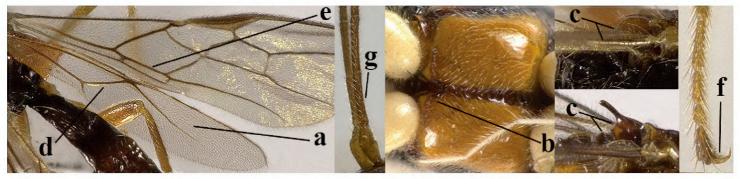




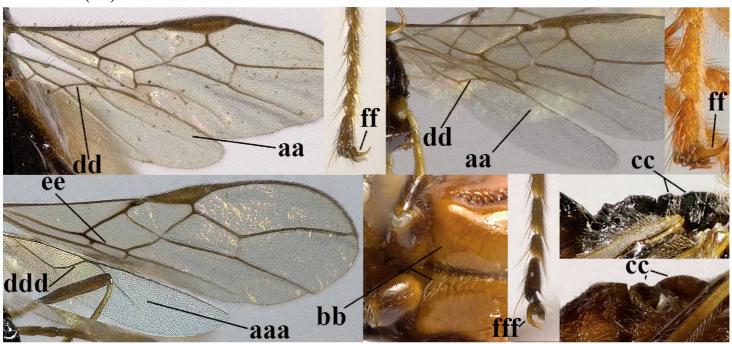








Marginal cell of hind wing slightly widened to narrowed (aa); **if** distinctly widened (aaa) then postpectal carina absent medio-ventrally (bb) and scutellum medio-posteriorly slightly or not protruding above level of metanotum in lateral view (cc); vein 1r-m of hind wing medium-sized, shorter than half width of hind wing (dd), **if** rarely longer (ddd) then first subdiscal cell of fore wing wider and shorter compared to vein cu-a (ee) and basal half of tarsal claws distinctly widened, subtriangular (ff), but sometimes parallel-sided (fff).

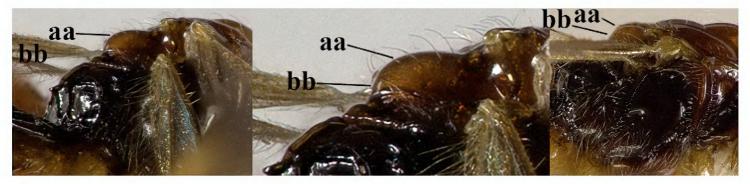


Scutellum of \mathcal{D} with distinct apical spine posteriorly (a), but sometimes less developed in \mathcal{D} ; scutellum steep medio-posteriorly in lateral view (b)

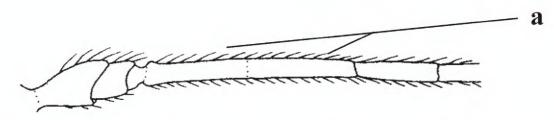
.....subgenus *Heratemis* Walker, 1860



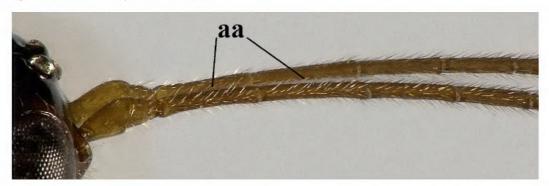
Scutellum of ♀ only distinctly convex posteriorly and without trace of a spine
 (aa); scutellum medio-posteriorly gradually lowered in lateral view (bb)....13

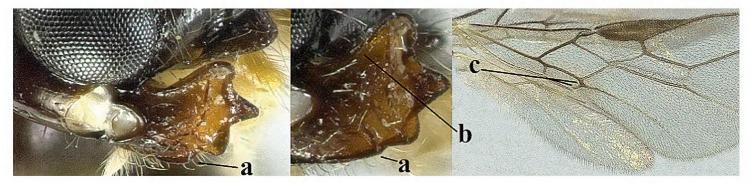


"Third" (actually joined third and fourth segments, sometimes vaguely separated) antennal segment 2.1–2.9 times as long as following segment and 9–11 times as long as wide (a).....subgenus *Kritscherysia* Fischer, 1993

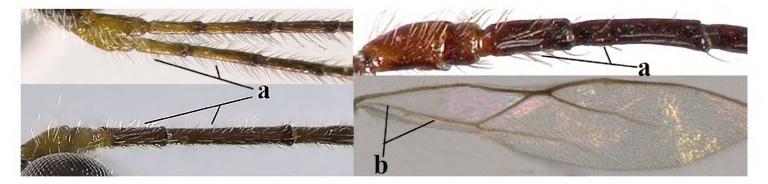


Third antennal segment 0.8–1.2 times following (= real fourth) segment and 4–7 times as long as wide (aa), **if** rarely third segment only partly separated from fourth segment, then its separation remains visible in lateral view subgenus Conalysia Papp, 1969

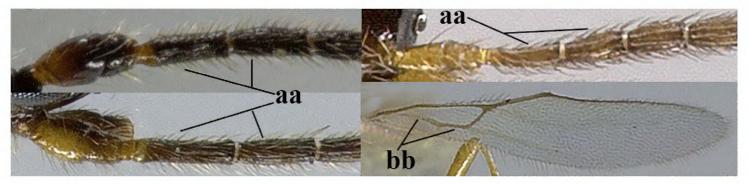


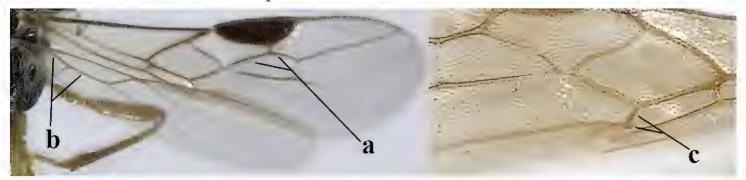




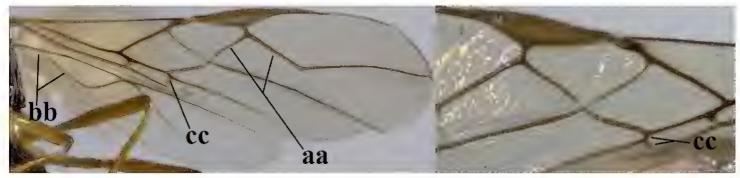


Third antennal segment subequal to or longer than fourth segment (aa); **if** subequal then vein M+CU of hind wing longer than vein 1-M (bb).......20





Vein 3-SR of fore wing longer than vein 2-SR (aa); **if** subequal then vein M+CU of hind wing distinctly shorter than vein 1-M (bb); vein CU1b of fore wing longer than vein 3-CU1 (cc); *Phaenocarpa* Foerster, 186317



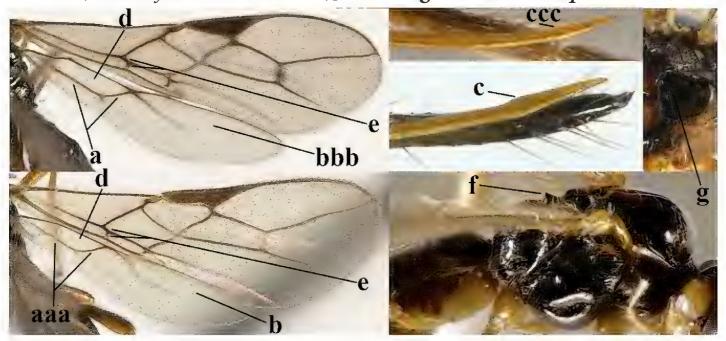
Tarsal claws distinctly widened medially and densely setose (especially swollen in ♀ and with apical tooth indistinct or small (a); but tarsal claws in ♂ slenderer and with distinct apical tooth, but still wider and more setose than in other groups) and pulvillus of ♀ strongly swollen; notauli complete, deep and crenulate (b).....subgenus *Discphaenocarpa* Belokobylskij, 1998

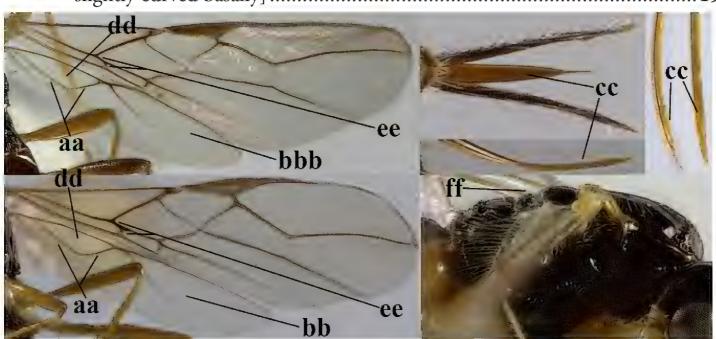


Tarsal claws flattened and with large apical tooth (aa) and pulvillus of ♀ not swollen; notauli often absent or smooth and shallow posteriorly (bb)18



Vein 1r-m of hind wing 0.2–0.7 times as long as vein 1-M (a); **if** 0.6–0.7 times (aaa) then metanotum tooth-shaped protruding dorsally in lateral view (f); marginal cell of hind wing medium-sized (bbb) or small (b); upper valve of ovipositor cylindrical and more or less widened subapically in lateral view (c), but in *P. ruficeps* group of equal width (ccc); apical half of basal cell of hind wing at most weakly widened (d); vein 1-CU1 of fore wing usually about as long as vein cu-a or shorter (e); [vein 1-SR+M of fore wing straight or slightly sinuate basally; vein 1-R1 of fore wing at least 1.6 times as long as pterostigma; metanotum tooth-shaped protruding in lateral view, vein 1r-m of hind wing 0.6–0.7 times as long as vein 1-M (0.2–0.5 times in other spp.) and the scutellar sulcus more or less narrowed medially in the *P. ruficeps* group (= *Holcalysia* Cameron, 1905)]....... **subgenus** *Phaenocarpa* **Foerster, 1863**



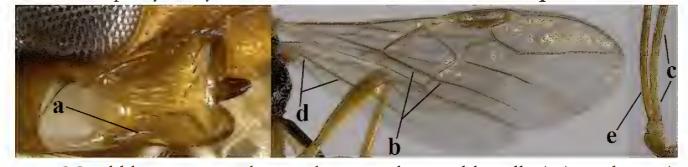


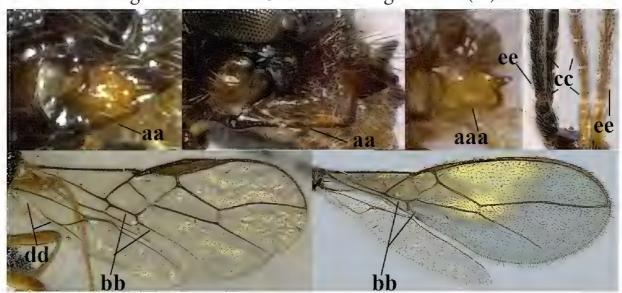
Vein 1-M of hind wing 0.8–1.2 times longer than vein M+CU (a); apically upper valve of ovipositor enclosed by much wider lower valve (b)......

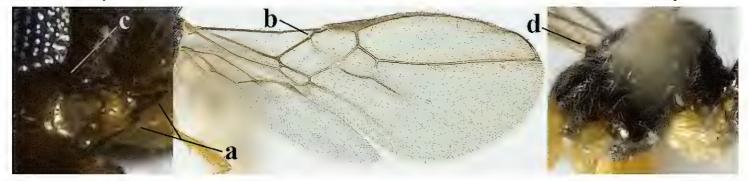
......subgenus Clistalysia Zhu, van Achterberg & Chen, 2017

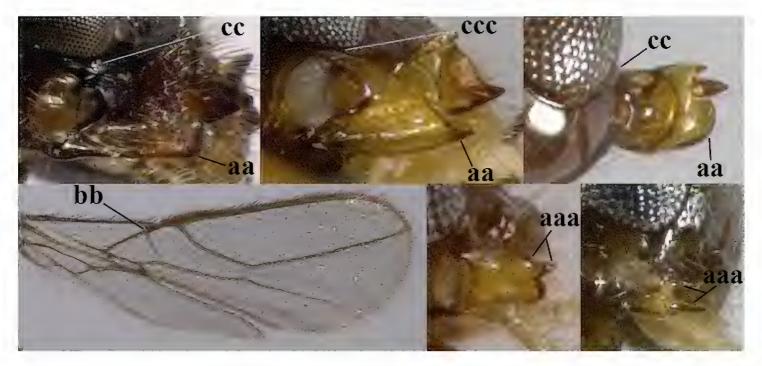


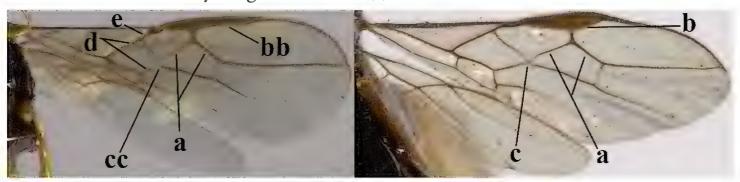


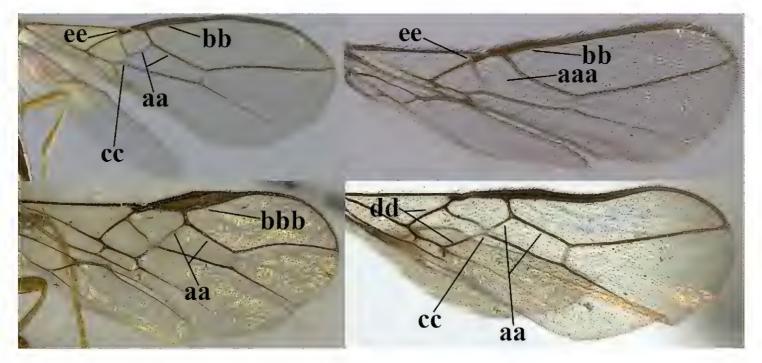


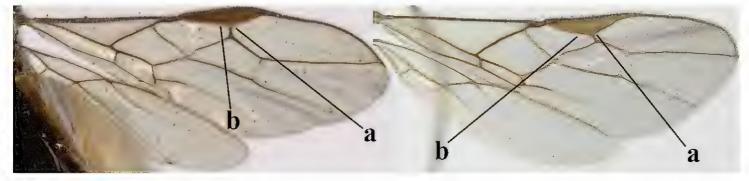




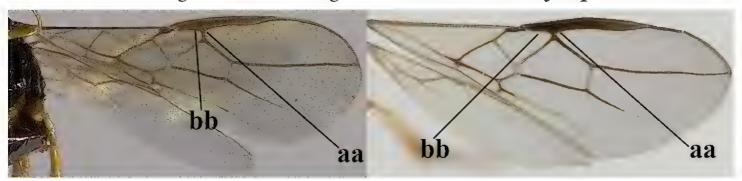


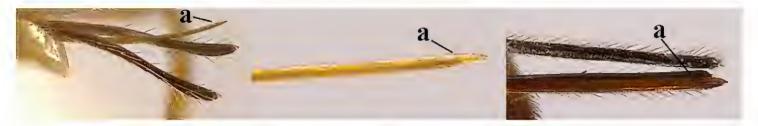


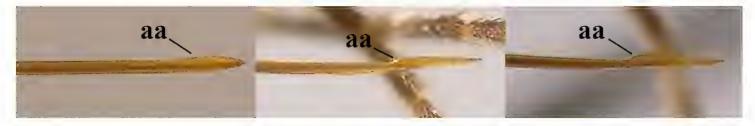


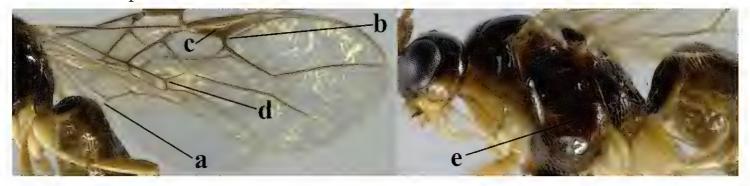


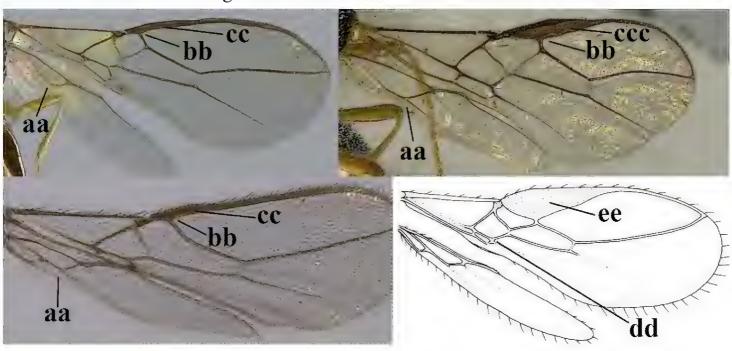
Vein r issued between basal third and middle of pterostigma (aa); pterostigma usually slender (bb); [temple posteriorly setose; tarsal claws often very slender submedially; second—fourth tarsal segments with long spines apically; apex of hind tibia with distinct whitish comb at inner side, but rarely absent; vein m-cu of fore wing about half as long as vein 1-M]................. *Tanycarpa* Foerster, 1863

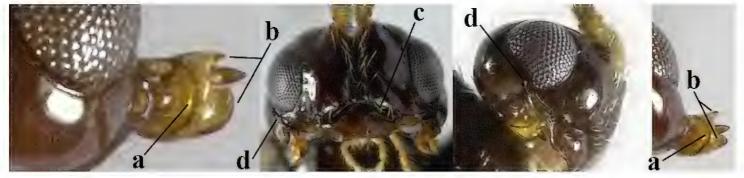


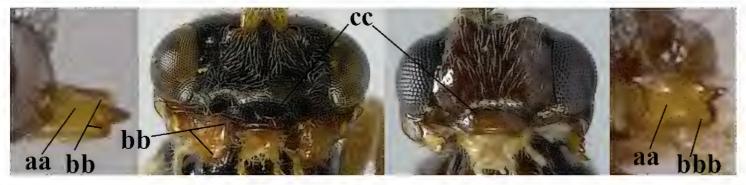






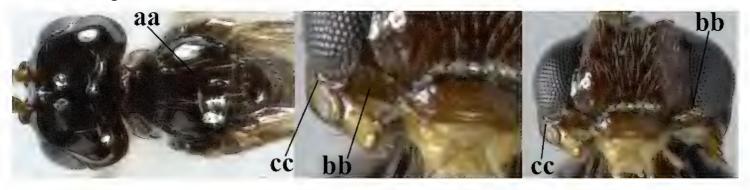


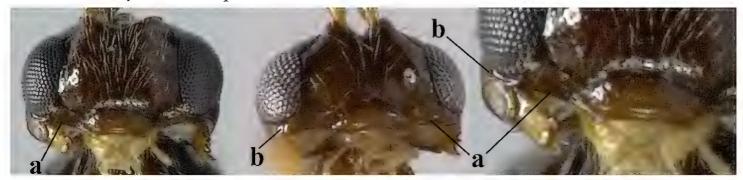






Notauli absent posteriorly, at most anterior half impressed (aa); anterior tentorial pit variable, if enlarged and flat (bb) then without an oblique subocular depression (cc)





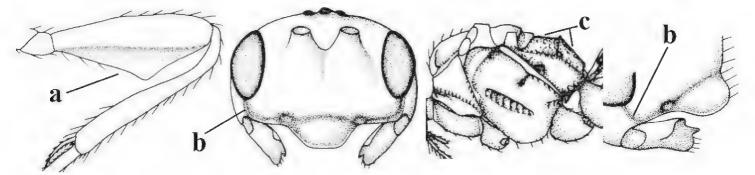


Vein 2-SR of fore wing present (a), but sometimes hardly sclerotized (aaa) ... subgenus *Aspilota* Foerster, 1863



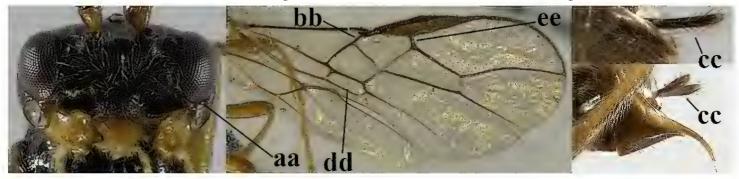


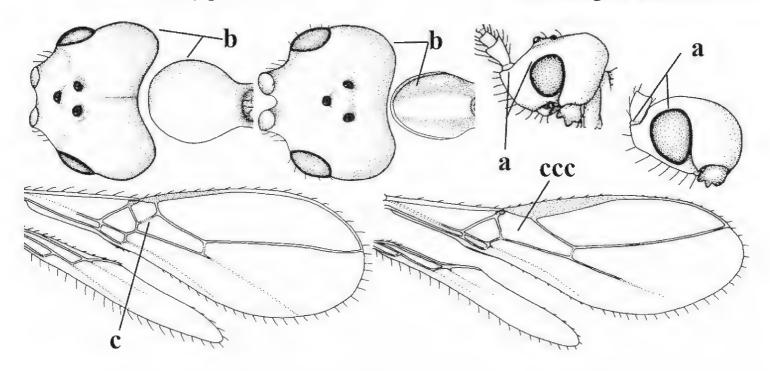
Fore femur with large obtuse tooth (flange) ventrally (a) or with row of minute teeth; malar suture subvertical (b); anterior part of propodeum differentiated and nearly as long as posterior part (c)......Leptotrema van Achterberg, 1988





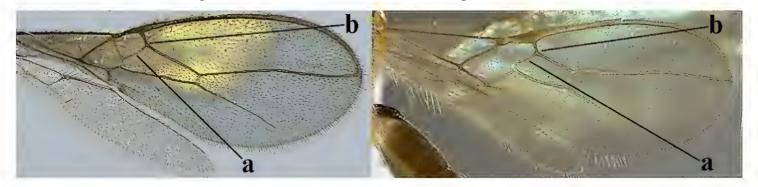


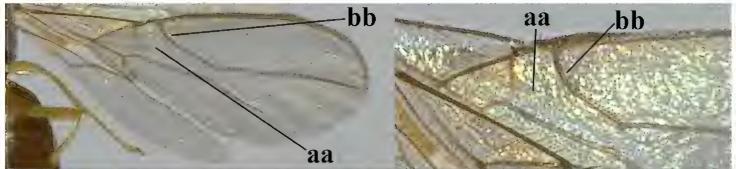


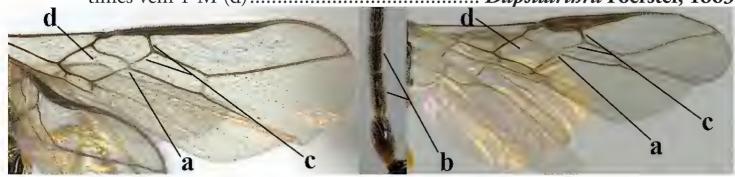




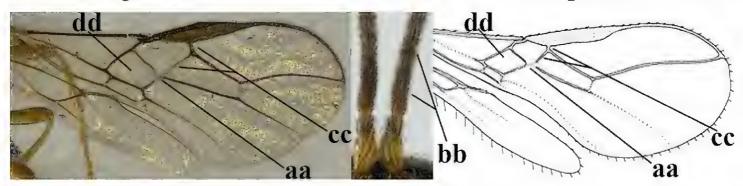
Vein 2-SR of fore wing present (a), **if** sometimes weakly sclerotised then vein r distinctly angled with vein 3-SR (b).....**subgenus** *Dinotrema* **Foerster**, **1863**







Vein m-cu of fore wing just antefurcal (aa); third antennal segment 1.2–1.5 times as long as fourth segment in Palaearctic spp. (bb); length of vein r of fore wing 0.2–0.3 times vein 2-SR (cc); diagonal width of first discal cell of fore wing often 1.4–1.6 times vein 1-M (dd) *Grammospila* Foerster, 1863



List of genera and species of Chinese Alysiini

Adelurola Strand, 1928

Adelurola Strand, 1928: 51 (nom. n. for Adelura Foerster, 1863); Shenefelt 1974: 986–987. Type species: Alysia florimela Haliday, 1838 (monobasic)

Synonym. Adelura Foerster, 1863, not Bonaparte, 1854; Neocarpa Fischer, 1966. **Biology.** Small genus, containing parasitoids of Tephritidae and Anthomyiidae. **Species.** Adelurola florimela Haliday, 1838.

Notes. Adelurola eurys Chen & Wu, 1994, belongs to Grammospila (comb. n.); it was transferred to Dapsilarthra Foerster by Peris-Filipo et al. (2016) because Dapsilarthra was used in a wider sense including Grammospila Foerster.

Alloea Haliday, 1833

Alloea Haliday, 1833: 265; Shenefelt 1974: 939; Chen and Wu 1994: 20; Belokobylskij 1998: 287. Type species: Alysia contracta Haliday, 1833.

Synonym. Diaspasta Foerster, 1863; Lamadatha Cameron, 1900.

Biology. Small genus, containing parasitoids of Lonchopteridae.

Species. Alloea ampla Wharton & Chou, 1985: Chen and Wu 1994

Alloea artus Chen & Wu, 1994

Alloea lineata Wharton & Chou, 1985: Chen and Wu 1994

Alloea lonchopterae Fischer, 1966: Chen and Wu 1994

Alloea mesostenos Chen & Wu, 1994

Alloea sparsa Wharton & Chou, 1985: Chen and Wu 1994

Alloea striata Wharton & Chou, 1985: Chen and Wu 1994

Alysia Latreille, 1804

Alysia Latreille, 1804: 173; Shenefelt 1974: 939; Wharton 1980a: 458; Chen and Wu 1994: 28; Belokobylskij 1998: 170. Type species: *Ichneumon manducator* Panzer, 1799.

Synonym. Cechenus Illiger, 1807; Anarcha Foerster, 1863 (subgenus); Goniarcha Foerster, 1863; Strophaea Foerster, 1863.

Biology. Large genus, containing parasitoids of Calliphoridae, Sarcophagidae, Tephritidae, Anthomyiidae, Agromyzidae and Mycetophylidae.

Notes. Typical species have vein m-cu of fore wing long (approx. 0.8 times 1-M) and 1-SR of fore wing linear with 1-M.

Species. Alysia (Alysia) frigida Haliday, 1838 (Chen and Wu 1994)

Alysia (Alysia) macrops Wharton, 1986 (Chen and Wu 1994)

Alysia (Alysia) manducator (Panzer, 1799) (Chen and Wu 1994)

Alysia (Anarcha) masneri Wharton, 1988 (Chen and Wu 1994)

Alysia (Alysia) nigritarsis Thomson, 1895 (Chen and Wu 1994)

Aphaereta Foerster, 1863

Aphaereta Foerster, 1863: 264: Shenefelt 1974: 956; Wharton 1980: 74; Chen and Wu 1994: 37; Belokobylskij 1998: 273; van Achterberg 2012: 2. Type species: Alysia cephalotes Haliday, 1833.

Biology. Rather small genus containing parasitoids of Agromyzidae, Anthomyiidae, Aulacigastridae, Calliphoridae, Chloropidae, Coelopidae, Fannidae, Muscidae, Ottidae, Sarcophagidae, Sciomyzidae, Tachinidae and Tephritidae.

Species. Aphaereta major (Thomson, 1895) (Chen and Wu 1994)

Aphaereta rubicunda Tobias, 1962 (Chen and Wu 1994)

Aphaereta scaptomyzae Fischer, 1966a (He and Chen 2004)

Aphaereta tricolor Papp, 1994 (He and Chen 2004)

Asobara Foerster, 1863

Asobara Foerster, 1863: 267; Shenefelt 1974: 964; Wharton 1980: 31; Chen and Wu 1994: 39; Belokobylskij 1998: 268; Wharton 2002: 28. Type species: Alysia tabida Nees von Esenbeck, 1834.

Synonym. Spanista Foerster, 1863.

Biology. Rather large genus, contains parasitoids of Drosophilidae and Sepsidae in decaying organic matter, especially fruits and leaves. The group with widened ovipositor sheath contains parasitoids of Tephritidae in fruits.

Species. Asobara aurea (Papp, 1967) (Papp 1967; Chou 1981; Chen and Wu 1994)

Asobara bactrocerae (Gahan, 1925) (Chen and Wu 1994)

Asobara elongata van Achterberg & Guerrieri, 2016 (Guerrieri et al. 2016)

Asobara formosae (Ashmead, 1906) (Fischer 1973a; Chou 1981; Ashmead 1906)

Asobara fungicola (Ashmead, 1894) (Chen and Wu 1994)

Asobara leveri (Nixon, 1939) (Chen and Wu 1994)

Asobara mesocauda van Achterberg & Guerrieri, 2016 (Guerrieri et al. 2016)

Asobara obliqua (Papp, 1969) (Chen and Wu 1994)

Asobara pleuralis (Ashmead, 1905) (Papp 1967; Guerrieri et al. 2016)

Asobara triangulata van Achterberg & Guerrieri, 2016 (Guerrieri et al. 2016)

Asobara tabida (Nees, 1834) (Chen and Wu 1994)

Asobara tabidula (Tobias, 1962) (Chen and Wu 1994)

Asobara unicolorata van Achterberg & Guerrieri, 2016 (Guerrieri et al. 2016)

Aspilota Foerster, 1863 s. s.

Aspilota Foerster, 1863: 268; Shenefelt 1974: 966; Wharton 1980: 84; van Achterberg 1988b: 9; Chen and Wu 1994: 49; Belokobylskij 1998: 218; Wharton 2002: 34. Type species: Alysia ruficornis Nees von Esenbeck, 1834 (monobasic).

Synonym. Dipiesta Foerster, 1863; Eusynaldis Zaykov & Fischer, 1982 (retained as subgenus with Regetus Papp, 1999 (syn. n.) and Adelphenaldis Fischer, 2003 (syn. n.) and Synaldis auctt. p.p. as synonyms).

Biology. Large genus, containing parasitoids of Phoridae and Platypezidae (in mushrooms). The host records of Anthomyiidae and Drosophilidae are probably erroneous.

Species. Aspilota (Eusynaldis) acutidentata (Fischer, 1970a) (Chen and Wu 1994)

Aspilota (Aspilota) elongata Chen & Wu, 1994 (Chen and Wu 1994)

Aspilota (Eusynaldis) globipes (Fischer, 1962) (Chen and Wu 1994)

Aspilota (Aspilota) intermediana Fischer, 1975 (Chen and Wu 1994)

Aspilota (Aspilota) louiseae van Achterberg, 1988 (Chen and Wu 1994)

Aspilota (Aspilota) nasica Belokobylskij, 2005 (Belokobylskij 2005; Belokobylskij and Tobias 2007)

Aspilota (Eusynaldis) parvicornis (Thomson, 1895) (Chen and Wu 1994)

Aspilota (Aspilota) schrenki Belokobylskij, 2007 (Belokobylskij and Tobias 2007)

Aspilota (Aspilota) tianmushanica Belokobylskij, 2005 (Belokobylskij 2005; Belokobylskij and Tobias 2007)

Aspilota (Aspilota) xuexini Belokobylskij, 2007 (Belokobylskij and Tobias 2007)

Notes. The genera *Regetus* Papp and *Adelphenaldis* Fischer share with *Eusynaldis* Zaykov & Fischer the derived character of the reduced vein 1-SR+M of the fore wing. The only difference between *Eusynaldis* and both other taxa is the shortened vein r-m of fore wing, a feature often variable within species of *Aspilota* Foerster and not suitable for separation of genera; the same applies to the enlarged propodeal spiracle of *Regetus* Papp. *Eusynaldis* Zaykov & Fischer is recognised as subgenus for convenience, because the recognition as genus likely renders the genus *Aspilota* Foerster paraphyletic, and the loss of vein 1-SR+M occurred probably more than once in the genus.

Carinthilota Fischer, 1975

Carinthilota Fischer, 1975: 311; van Achterberg 1988b: 17; Chen and Wu 1994: 59; Belokobylskij 1998: 221. Type species: Carinthilota parapsidalis Fischer, 1975.

Biology. Unknown, but related genera have been reared from Phoridae and Platypezidae. **Species.** *Carinthilota parapsidalis* Fischer, 1975 (Chen and Wu 1994)

Cratospila Foerster, 1863

Cratospila Foerster, 1863: 265; Shenefelt 1974: 985; Wharton 1980: 84; Tobias 1990; Belokobylskij 1998: 287; Yao 2016: 1. Type species: Alysia circe Haliday, 1838.

Synonym. Hedylus Marshall, 1894 (not Foerster 1868).

Biology. Rather small genus, of which the biology is unknown.

Species. Cratospila circe (Haliday, 1838) (Wu and Chen 1995a)

Dacnulysia Zhu, van Achterberg & Chen, 2017

Dacnulysia Zhu, van Achterberg & Chen, 2017: 361.

Biology. Unknown.

Species. Dacnulysia chaenomastax Zhu, van Achterberg & Chen, 2017

Dapsilarthra Foerster, 1863

Dapsilarthra Foerster, 1863: 267. Shenefelt 1974: 986–991; Marsh 1979: 222; Wharton 1980: 37–38; van Achterberg 1983a: 6–14; Chen and Wu 1994: 61; Belokobylskij 1998: 208–209. Type species: *Alysia apii* Curtis, 1826 (monobasic).

Biology. Small genus, containing parasitoids of Agromyzidae. **Species.** Dapsilarthra apii (Curtis, 1826) (Chen and Wu 1994) Dapsilarthra sylvia (Haliday, 1839) (Chen and Wu 1994)

Dinotrema Foerster, 1863

Dinotrema Foerster, 1863: 268; Shenefelt 1974: 966; Wharton 1980: 84; van Achterberg and Bin 1981: 104; Chen and Wu 1994: 69; Wharton 2002: 56; Tobias 2003: 138. Type species: Dinotrema erythropa Foerster, 1863 (monobasic).

Synonym. Spanomeris Foerster, 1863; Coloboma Foerster, 1863; Prosapha Foerster, 1863; Synaldis Foerster, 1863 (subgenus); Synaldotrema Belokobylskij & Tobias, 2007 (subgenus); Aspilota auctt. p. p.

Biology. Very large genus, containing parasitoids of Phoridae.

Species. Dinotrema (Dinotrema) amoenidens (Fischer, 1973b) (Chen and Wu 1994)

Dinotrema (Dinotrema) cato Tobias, 2007 (Belokobylskij and Tobias 2007)

Dinotrema (Dinotrema) conjunctum Tobias, 2007 (Belokobylskij and Tobias 2007)

Dinotrema (Synaldis) distractum (Nees, 1834) (Chen and Wu 1994)

Dinotrema (Dinotrema) hodisense (Fischer, 1976) (Chen and Wu 1994)

Dinotrema (Dinotrema) kempei (Hedqvist, 1973) (Chen and Wu 1994)

Dinotrema (Dinotrema) longus (Wu & Chen, 1998) (Wu and Chen 1998a)

Dinotrema (Synaldis) mandibulatum (Fischer, 1970) (Chen and Wu 1994)

Dinotrema (Dinotrema) mesocaudatum van Achterberg, 1988 (Chen and Wu 1994)

Dinotrema (Dinotrema) monstrconnexum Tobias, 2007 (Belokobylskij and Tobias 2007)

Dinotrema (Dinotrema) multiarticulatum van Achterberg, 1988 (Chen and Wu 1994)

Dinotrema (Dinotrema) nitidula (Masi, 1933) (Chen and Wu 1994)

Dinotrema (Dinotrema) occipitale (Fischer, 1973) (Chen and Wu 1994)

Dinotrema (Dinotrema) pratense van Achterberg, 1988 (Chen and Wu 1994)

Dinotrema (Dinotrema) pulvinatum (Stelfox & Graham, 1949) (Chen and Wu 1994)

Dinotrema (Dinotrema) tauricum (Telenga, 1935) (Chen and Wu 1994)

Dinotrema (Dinotrema) tuberculatum van Achterberg, 1988 (Chen and Wu 1994)

Notes. A diverse genus including several spp. without oblique subocular depression for which the names *Prosapha* Foerster, 1863, *Panerema* Foerster, 1863, and *Pterusa* Fischer, 1958, are available. An extensive worldwide phylogenetic study of the genus *Dinotrema* is necessary before a well-based decision can be made on a possible recognition as subgenus or genus. *Synaldis* Foerster is recognised as subgenus for convenience, because the recognition as genus likely renders the genus *Dinotrema* Foerster paraphyletic, and the loss of vein 1-SR+M occurred probably more than once in the genus.

Eudinostigma Tobias, 1986

Eudinostigma Tobias, 1986: 244; Chen and Wu 1994: 78; Belokobylskij 1998: 219. Type species: Eudinostigma fischeri Tobias, 1986.

Synonym. According to Wharton (2002) a synonym of *Dinotrema* Foerster, 1863.

Biology. Small genus, of which the biology is unknown, but related species are parasitoids of Phoridae.

Species. Eudinostigma alox van Achterberg, 1988 (Chen and Wu 1994)

Eudinostigma latistigma (Fischer, 1962) (Wu and Chen 1998b)

Eudinostigma latus Chen & Wu, 1994. (Chen and Wu 1994)

Grammospila Foerster, 1863

Grammospila Foerster, 1863: 269; Shenefelt 1974: 987; van Achterberg 1983a: 7. Type species: Alysia isabella Haliday, 1838 (monobasic).

Synonym. Paraorthostigma Königsmann, 1972.

Biology. Small genus, containing parasitoids of Agromyzidae and Scathophagidae.

Species. Grammospila eurys (Chen & Wu, 1994), comb. n.

Grammospila isabella (Haliday, 1838) (Chen and Wu 1994)

Grammospila rufiventris (Nees, 1812) (Chen and Wu 1994)

Notes. Grammospila eurys (Chen & Wu, 1994), comb. n. has the third antennal segment 1.4–1.5 times as long as fourth segment; vein m-cu of fore wing antefurcal (not post-furcal as mentioned in original (Chinese) description); body with many long setae (includ-

ing mesoscutum); vein r of fore wing widened, hardly longer than wide; base of pterostigma slender and posteriorly concave and pterostigma up to level of vein r-m of fore wing.

Heratemis Walker, 1860

Heratemis Walker, 1860: 310; Fischer 1966b: 177; Shenefelt 1974: 992; Chen and Wu 1994: 82; Belokobylskij 1998: 268; Wharton 2002: 75; Yaakop et al. 2009: 1. Type species: Heratemis filosa Walker, 1860 (monobasic).

Synonym. Conalysia Papp, 1969 (subgenus); Kritscherysia Fischer, 1993 (subgenus).

Biology. Medium-sized genus, of which the biology is unknown, possibly parasitoids of Tephritidae.

Species. Heratemis (Conalysia) devriesi van Achterberg & Yaakop, 2009 (Yaakop et al. 2009)

Heratemis (Kritscherysia) enodis Wu & Chen, 1994 (Chen and Wu 1994)

Heratemis (Heratemis) filosa Walker, 1860 (Chen and Wu 1994; Yaakop et al. 2009)

Heratemis (Conalysia) laticeps (Papp, 1969) (Chen and Wu 1994; Yaakop et al. 2009)

Heratemis (Conalysia) ustulata Wu & Chen, 1996 (Wu and Chen 1996)

Notes. Morphologically *Heratemis* spp. are very similar to species of the subgenus *Neophaenocarpa* Belokobylskij of the genus *Phaenocarpa* Foerster. The presence of the postpectal carina and the posteriorly steep scutellum of *Heratemis* allow a clear separation.

Heterolexis Foerster, 1863

Heterolexis Foerster, 1863: 268; Shenefelt 1974: 992; van Achterberg 1983a: 7. Type species: Heterolexis subtilis Foerster, 1863.

Biology. Small genus, containing parasitoids of Agromyzidae and Anthomyiidae. **Species.** *Heterolexis subtilis* Foerster, 1863 (Chen and Wu 1994)

Hylcalosia Fischer, 1967

Hylcalosia Fischer, 1967: 125; Shenefelt 1974: 993; Chen and Wu 1994: 85; Belokobylskij 1998: 297; Zheng et al. 2012: 454. Type species: Holcalysia testaceipes Cameron, 1910.

Synonym. Holcalysia Cameron, 1910, not Cameron 1905.

Biology. Small genus, of which the biology is unknown.

Species. Hylcalosia complexa Chen & Wu, 1994 (Chen and Wu 1994; Zheng et al. 2012)

Hylcalosia ventisulcata Zheng, Chen & Yang, 2012 (Zheng et al. 2012)

Idiasta Foerster, 1863

Idiasta Foerster, 1863, 265; Shenefelt 1974: 993; Chen and Wu 1994: 87; Belokobylskij 1998: 277. Type species: *Alysia maritima* Haliday, 1838.

Synonym. Euphaenocarpa Tobias, 1975.

Biology. Medium-sized genus, containing parasitoids of Muscidae.

Species. Idiasta annulicornis (Thomson, 1895) (Chen and Wu 1994)

Idiasta brevicauda Telenga, 1935 (Chen and Wu 1994)

Idiasta dichrocera Königsmann, 1960 (Chen and Wu 1994)

Idiasta paramaritima Königsmann, 1960 (Chen and Wu 1994)

Idiasta picticornis (Ruthe, 1854) (Chen and Wu 1994)

Idiasta subannellata (Thomson, 1895) (Chen and Wu 1994)

Leptotrema van Achterberg, 1988

Leptotrema van Achterberg, 1988a: 42; Chen and Wu 1994: 94; Belokobylskij 1998: 219. Type species: Aspilota dentifemur Stelfox, 1943.

Synonym. According to Wharton (2002) this is a synonym of *Dinotrema* Foerster, 1863. However, the vertical malar suture excludes it from *Dinotrema* Foerster. A future DNA-analysis is needed to find its position within the *Aspilota*-group.

Biology. Small genus of which the biology is unknown, but belongs to the *Aspilota*-group containing parasitoids of Phoridae.

Species. Leptotrema dentifemur (Stelfox, 1943) (Chen and Wu 1994)

Mesocrina Foerster, 1863

Mesocrina Foerster, 1863: 266; Shenefelt 1974: 996; Chen and Wu 1994: 95; Beloko-bylskij 1998: 191. Type species: Mesocrina indagatrix Foerster, 1863.

Synonym. Pseudomesocrina Königsmann, 1959.

Biology. Small genus, containing parasitoids of Anthomyiidae and Scathophagidae, the type species is associated with hosts in mushrooms.

Species. Mesocrina dalhousiensis (Sharma, 1978) (Chen and Wu 1994) Mesocrina indagatrix Foerster, 1863 (Chen and Wu 1994) Mesocrina licho Belokobylskij, 1998 (new to China)

Mesocrina licho Belokobylskij, 1998

Figs 1–14

Material. ♀ (ZJUH), "[N. China:], Hebei, Mt. Xioawutai, 23.viii.2005, Shi Min, No. 200608887"; 2 ♂♂ (ZJUH), id., but Zhang Hongying, No. 200609036, 200609050; 2 ♂♂ (ZJUH), id., but 21.viii.2005, Zhang Hongying, 200608013, 200608045.

Description of ♀ **from Mt. Xioawutai.** Length of body 3.9 mm, of fore wing 4.6 mm. *Head.* Transverse and shiny (Fig. 9), width of head twice its lateral length; antenna incomplete, with 23 remaining segments, segments with bristly setae, third segment 1.4 times longer than fourth segment, length of third and fourth segments 5.0 and 3.8 times their width, respectively (Fig. 7); length of maxillary palp twice height of head; eye in dorsal view 1.4 times as long as temple (Fig. 9); eye in lateral view1.4 times higher than wide; vertex convex and glabrous (Fig. 11); OOL:diameter of ocellus:POL= 9:5:5; face 1.7 times wider than high, smooth and shiny (Fig. 10), with some long setae next to eye; clypeus medium-sized, rather flat, truncate and slightly convex laterally (Fig. 10); malar space absent; mandible moderately widened dorsally, dorsal teeth large and lobe-shaped (Fig. 12), lateral teeth rather small and lobe-shaped (Fig. 13), middle tooth curved and acute; medial length of mandible 1.5 times its maximum width (Fig. 13).

Mesosoma. Length of mesosoma 1.3 times its height; mesoscutum without lateral carina in front of tegula (Fig. 3); precoxal sulcus absent; mesopleuron smooth and glabrous; pleural sulcus crenulate; episternal scrobe small, connected by a furrow to pleural sulcus; metapleuron smooth except some ventral rugae, with long setae and a round large pit anteriorly (Fig. 3); notauli only anteriorly impressed on disc, narrowly crenulate and medio-posteriorly with deep longitudinal depression; mesoscutum with some setae anteriorly and near notauli; scutellar sulcus deep and narrow, with 4 short longitudinal carinae and 6 times wider than its maximum length; scutellum rather flat and wide (Fig. 4); surface of propodeum with rather long median carina, without areola absent and with some rugae anteriorly (Fig. 5).

Wings (Fig. 2). Pterostigma largely wide elliptical, vein r 0.5 times width of pterostigma; r:3-SR:SR1 = 5:33:67; SR1, 1-SR+M nearly straight and 2-SR slightly curved; cu-a postfurcal, short; 1-CU1:2-CU1 = 2:17; 3-CU1 longer than CU1b; 2-SR:3-SR:r-m = 19:25:8; m-cu postfurcal, slightly converging to 1-M posteriorly; first subdiscal cell 3.3 times as long as wide; M+CU1 largely unsclerotized. Hind wing: M+CU: 1-M:1r-m = 25:23:20; m-cu present.



Figure 1. *Mesocrina licho* Belokobylskij, ♀, China, Mt. Xioawutai, habitus lateral.

Legs. Hind coxa smooth; tarsal claws rather robust and longer than arolium (Fig. 1); length of femur, tibia and basitarsus of hind leg 4.3, 10.0 and 6.7 times their width, respectively; apical spiny bristles of first-fourth hind tarsal segments absent (Fig. 1).

Metasoma. Length of first tergite 1.3 times its apical width, its surface with longitudinal striae, its dorsal carinae narrowly connected (Fig. 5); laterope absent; dorsope rather large (Fig. 6); setose part of ovipositor sheath 0.18 times as long as fore wing (total visible sheath 0.19 times), flattened and sparsely setose and 0.6 times as long as hind tibia (Fig. 8).

Colour. Blackish brown (Fig. 1); pronotum ventrally, mandible, tegula, two basal segments of antenna, palpi mainly pale and remainder of legs yellowish; antenna (ex-



Figures 2–14. *Mesocrina licho* Belokobylskij, ♀, China, Mt. Xioawutai. 2 wings 3 mesosoma lateral 4 mesosoma dorsal 5 propodeum, first and second metasomal tergites dorsal 6 propodeum and metasoma dorsal 7 basal segments of antenna lateral 8 ovipositor and sheath lateral 9 head dorsal 10 head anterior 11 head lateral 12 full view of first and second tooth of mandible 13 full view of third tooth of mandible 14 antenna lateral.

cept two basal segments of antenna), head (except ventrally), mesosoma, dorsal spot of hind femur, hind tibia (except basally) and basitarsus, and first tergite of metasoma blackish brown; head ventrally, mesopleuron ventrally and remainder of metasoma brown; pterostigma and veins brown; wing membrane slightly infuscated.

Variation. Males are similar to females, but have 35(1) antennal segments (according to the original description females have 31 or 32 segments); body length of 3.7-4.2 mm, length of fore wing 4.1-4.7 mm, width of head 1.9-2.0 times its lateral length.

Orthostigma Ratzeburg, 1844

Orthostigma Ratzeburg, 1844: 53; Shenefelt 1974: 997; Wharton 1980: 85; van Achterberg 1988b: 44; Chen and Wu 1994: 99; Belokobylskij 1998: 209. Type species: Aphidius flavipes Ratzeburg, 1844.

Synonym. *Delocarpa* Foerster, 1863; *Ischnocarpa* Foerster, 1863; *Afrostigma* Fischer, 1995 (subgenus); *Patrisaspilota* Fischer, 1995 (subgenus).

Biology. Medium-sized genus, containing parasitoids of Phoridae. The records of Agromyzidae, Cecidomyiidae, and Drosophilidae are probably erroneous.

Species. Orthostigma cratospilum (Thomson, 1895) (Chen and Wu 1994)

Orthostigma imperator van Achterberg & Ortega, 1983 (Chen and Wu 1994)

Orthostigma laticeps (Thomson, 1895) (Chen and Wu 1994)

Orthostigma lokei Hedqvist, 1973 (Chen and Wu 1994)

Orthostigma longicorne Königsmann, 1969 (Chen and Wu 1994)

Orthostigma longicubitale Königsmann, 1969 (Chen and Wu 1994)

Orthostigma lucidum Königsmann, 1969 (Chen and Wu 1994)

Orthostigma mandibulare (Tobias, 1962) (Chen and Wu 1994)

Orthostigma pumilum (Nees, 1834) (Chen and Wu 1994)

Orthostigma pusillum (Zetterstedt, 1838) (Chen and Wu 1994)

Orthostigma sculpturatum Tobias, 1962 (Chen and Wu 1994)

Orthostigma sibiricum (Telenga, 1933) (Chen and Wu 1994)

Orthostigma sordipes (Thomson, 1895) (Chen and Wu 1994)

Phaenocarpa Foerster, 1863

Phaenocarpa Foerster, 1863: 267; Papp, 1968: 570; Fischer, 1970b: 409; Shenefelt, 1974: 1003; Wharton, 1980: 96; Chen & Wu, 1994: 114; Belokobylskij, 1998: 233. Type species: *Alysia picinervis* Haliday, 1838.

Synonym. Homophyla Foerster, 1863 (subgenus); Mesothesis Foerster, 1863; Sathra Foerster, 1863; Idiolexis Foerster, 1863 (subgenus); Asynaphes Provancher, 1886; Kahlia Ashmead, 1900 (subgenus); Stiralysia Cameron, 1910; Rhopaloneura Stelfox, 1941;

Discphaenocarpa Belokobylskij, 1998 (subgenus); Neophaenocarpa Belokobylskij, 1998 (subgenus); Sibphaenocarpa Belokobylskij, 1998 (subgenus); Uncphaenocarpa Belokobylskij, 1998 (subgenus); Ussurphaenocarpa Belokobylskij, 1998 (subgenus); Clistalysia Zhu, van Achterberg & Chen, 2017 (subgenus).

Biology. Large genus, containing koinobiont endoparasitoids of larvae of cyclor-rhaphous Diptera in many niches. Known from larvae of Sciomyzidae in Mollusca, of Syrphidae under bark or between leaves of marsh plants, of Anthomyiidae in roots of vegetables, under bark, in cones of conifers, mining in leaves or in dung, of Muscidae and Scathophagidae in dung, of Muscidae and Clusiidae in flood refuse and of Chloropidae and Scathophagidae in grasses and Drosophilidae in crops (e.g. cotton) and slime (Wharton, 1984; van Achterberg, 1998).

Species. Phaenocarpa (Phaenocarpa) cameroni Papp, 1967 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) carinthiaca Fischer, 1975 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) conspurcator (Haliday, 1838) (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) diffusa Chen & Wu, 1994 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) eunice (Haliday, 1838) (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) galatea (Haliday, 1838) (Wu and Chen 1995b) Phaenocarpa (Phaenocarpa) impressinotum Fischer, 1975 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) ingressor Marshall, 1896 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) intermedia Tobias, 1962 (Wu and Chen 1995b) Phaenocarpa (Phaenocarpa) laticellula Papp, 1968 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) lissogastra Tobias, 1986 (Belokobylskij 1998) Phaenocarpa (Phaenocarpa) notabilis Stelfox, 1944 (Chen and Wu 1994) Phaenocarpa (Clistalysia) platychora Zhu, van Achterberg & Chen, 2017 Phaenocarpa (Phaenocarpa) pratellae (Curtis, 1826) (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) riphaeica Tobias, 1986 (Wu and Chen 1995b) Phaenocarpa (Phaenocarpa) ruficeps (Nees, 1812) (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) seitneri Fahringer, 1929 (Chen and Wu 1994) Phaenocarpa (Phaenocarpa) vitata Chen & Wu, 1994 (Chen and Wu 1994)

Notes. Some species (e.g., *P. stackelbergi* Tobias & Gurasashvili, 1985) are superficially similar to *Idiasta* Foerster, because the \mathcal{P} antenna has a white band and the metanotum has an acute tooth in lateral view.

Separatatus Chen & Wu, 1994

Separatatus Chen & Wu, 1994: 132. Type species: Separatatus carinatus Chen & Wu, 1994.

Synonym. *Phasmidiasta* sensu Fischer, 2006, not Wharton 1980; *Hovalysia* sensu Wharton, 2002 (p. p.); *Bobekoides* auct. p. p.

Biology. Small genus, of which the biology is unknown.

Species. Separatatus carinatus Chen & Wu, 1994

Separatatus sinicus (Zheng, Chen & Yang, 2012), comb. n.

Separatatus parallelus sp. n.



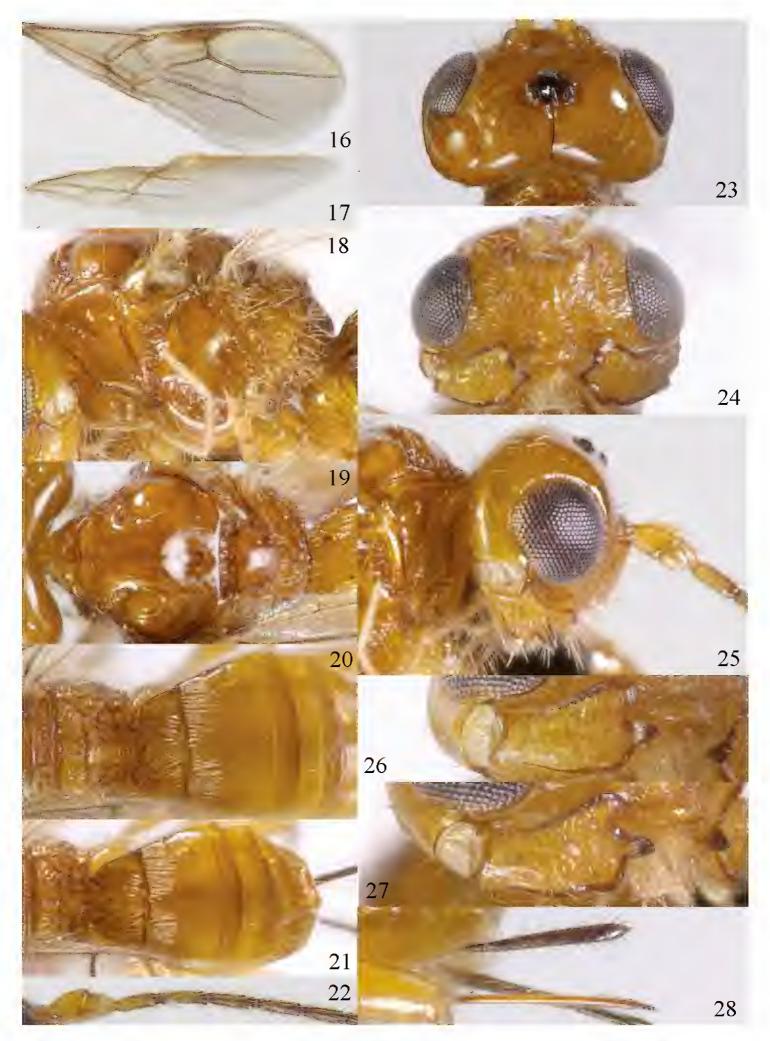
Figure 15. *Separatatus parallelus* sp. n., ♀, holotype, habitus lateral.

Separatatus parallelus sp. n.

http://zoobank.org/CB7FCC77-14F8-4080-8899-D23DA5A76D4E Figs 15–28

Material. Holotype, ♀ (ZJUH), "[S. China:], Yunnan, green water nuclear power station, 536 m, 23.vii.2003, Xu Zaifu, No. 20055387". Paratype: 1 ♂ (ZJUH), "Hainan, Yinggeling, 283.v.2007, Weng Liqiong, No. 200804310".

Description. Holotype, ♀, length of body 2.5 mm, of fore wing 2.6 mm.



Figures 16–28. Separatatus parallelus sp. n., ♀, holotype, **16** fore wing **17** hind wing **18**, mesosoma lateral **19** mesosoma dorsal **20** propodeum, first and second metasomal tergites dorsal **21** propodeum and metasoma lateral **22** basal segments of antenna **23** head dorsal **24**, head anterior **25** head lateral **26** mandible full view of first and second tooth mandible **27** mandible full view of third tooth **28** ovipositor and sheath lateral.

Head. Transverse and shiny, concave posteriorly (Fig. 23), width of head 1.8 times its lateral length; antenna incomplete, with 21 remaining segments, segments with bristly setae, third segment 0.7 times longer than fourth segment, length of third and fourth segments 2.5 and 4.7 times their width, respectively (Fig. 22); length of maxillary palp 1.4 times height of head; eye in dorsal view 2.1 times as long as temple (Fig. 23); eye in lateral view nearly as high as wide; vertex convex and glabrous (Fig. 25); OOL:diameter of ocellus:POL= 14:3:5; face 1.8 times wider than high, largely rugose (Fig. 24); clypeus rather small, truncate and slightly convex laterally (Fig. 24); malar space absent; mandible moderately widened dorsally, dorsal teeth large and lobeshaped (Fig. 26), lateral teeth rather small and lobe-shaped (Fig. 27), middle tooth curved; medial length of mandible 1.6 times its maximum width (Fig. 27).

Mesosoma. Length of mesosoma 1.4 times its height; mesoscutum without lateral carina in front of tegula (Fig. 18); epicnemial area smooth except for a few crenulae; precoxal sulcus wide, with distinct crenulae medially, but anteriorly and posteriorly absent; remainder of mesopleuron smooth and glabrous; pleural sulcus narrowly crenulate; episternal scrobe small, connected by a furrow to pleural sulcus; metapleuron reticulate-rugose but smooth medially, with long setae and a round large pit anteriorly (Fig. 18); notauli wide, only anteriorly impressed on disc, widely crenulate and medio-posteriorly with a shallow, round depression; mesoscutum with some setae along notauli; scutellar sulcus deep and narrow, with one median carina and 2 short longitudinal carinae and 4.0 times wider than its maximum length; scutellum rather flat and wide (Fig. 19); surface of propodeum rugose, with rather distinct median carina on anterior half, areola present but inconspicuous (Fig. 20).

Wings (Figs 16, 17). Pterostigma elliptical, vein r 0.8 times width of pterostigma; r:3-SR:SR1 = 5:14:40; SR1, 1-SR+M nearly straight and 2-SR curved; cu-a postfurcal, short; 1-CU1:2-CU1 = 2:17; 3-CU1 longer than CU1b; 2-SR:3-SR:r-m = 19:25:8; m-cu postfurcal, slightly converging to 1-M posteriorly; first subdiscal cell 3.8 times as long as wide; M+CU1 unsclerotised. Hind wing: M+CU: 1-M:1r-m = 4:3:2; m-cu absent.

Legs. Hind coxa smooth; tarsal claws rather robust and shorter than arolium (Fig. 15); length of femur, tibia and basitarsus of hind leg 2.7, 7.5 and 5.0 times their width, respectively; apical bristles of first-fourth hind tarsal segments absent (Fig. 15).

Metasoma. Length of first tergite 0.7 times its apical width, its surface longitudinally striate, its dorsal carinae widely separate (Fig. 20); second tergite of metasoma with longitudinally striate anteriorly; laterope present; dorsope rather large (Fig. 21); setose part of ovipositor sheath 0.26 times as long as fore wing (total visible sheath 0.35 times), flattened and sparsely setose and 0.8 times as long as hind tibia.

Colour. Yellowish brown (Fig. 15); palpi yellow; 4 basal segments of antenna, pterostigma and veins yellowish brown; wing membrane slightly infuscated.

Variation. Male is similar to female; body length of $3 \cdot 2.3$ mm, length of fore wing 2.4 mm, width of head 2.0 times its lateral length.

Notes. The new species can be separated from all known species by the parallel-sided and long basal part of the pterostigma, vein r of fore wing comparatively close to the apex of the pterostigma and vein 3-SR of fore wing about $2.9 \times as$ long as vein r.

Tanycarpa Foerster, 1863

Tanycarpa Foerster, 1863: 26; Chen and Wu 1994: 133; Belokobylskij 1998: 198; Yao 2015a: 170. Type species: *Bassus gracilicornis* Nees von Esenbeck, 1812 (monobasic and original designation).

Synonym. Acrobela Foerster, 1863; Epiclista Foerster, 1863.

Biology. Small genus, containing parasitoids primarily of Drosophilidae and Mycetophilidae in rotting plant or fungal substrates.

Species. Tanycarpa amplipennis (Foerster, 1863) (Chen and Wu 1994; Yao 2015a).

Tanycarpa areolata Yao, 2015 (Yao 2015a).

Tanycarpa bicolor (Nees, 1812) (Chen and Wu 1994; Yao 2015a).

Tanycarpa chors Belokobylskij, 1998 (Yao 2015a).

Tanycarpa concreta Chen & Wu, 1994 (Chen and Wu 1994; Yao 2015a).

Tanycarpa gladia Chen & Wu, 1994 (Chen and Wu 1994; Yao 2015a).

Tanycarpa gracilicornis (Nees, 1812) (Chen and Wu 1994; Yao 2015a).

Tanycarpa gymnonotum Yao, 2015 (Yao 2015a).

Tanycarpa lineata Yao, 2015 (Yao 2015a).

Tanycarpa mitis Stelfox, 1941 (Chen and Wu 1994; Yao 2015a).

Tanycarpa punctata van Achterberg, 1976 (Chen and Wu 1994; Yao 2015a).

Tanycarpa rufinotata (Haliday, 1838) (Chen and Wu 1994; Yao 2015a).

Tanycarpa scabrator Chen & Wu, 1994 (Chen and Wu 1994; Yao 2015a).

Tanycarpa similis Yao, 2015 (Yao 2015a)

Trachyusa Ruthe, 1854

Trachyusa Ruthe, 1854: 351; Yao 2015b: 580. Type species: Trachyusa nigriceps Ruthe, 1854.

Synonym. Cosmiocarpa Foerster, 1863.

Biology. Small genus, of which the biology is unknown. The record of Cimbicidae is erroneous.

Species. Trachyusa whartoni Yao, 2015 (Yao 2015b).

Acknowledgements

We are grateful to Dr Dicky Yu (Nepean) for providing many references. Funding for this study was provided by the State Key Program of National Natural Science Foundation of China (31230068) and the 973 Program (2013CB127600).

References

- Ashmead WH (1894) Descriptions of thirteen new parasitic Hymenoptera, bred by Prof. F.M. Webster. Journal of the Cincinnati Society of Natural History 17: 45–55.
- Ashmead WH (1900) Classification of the Ichneumon flies, or the superfamily Ichneumonoidea. Proceedings of the United States National Museum 23(1206): 1–220. https://doi.org/10.5479/si.00963801.23-1206.1
- Ashmead WH (1906) Despriptions of new Hymenoptera from Japan. Proceedings of the United States National Museum 30: 169–201. https://doi.org/10.5479/si.00963801.30-1448.169
- Ashmead WH (1905) New Hymenoptera from the Philippine Islands. Canadian Entomologist 37(1): 3–8. https://doi.org/10.4039/Ent373-1
- Belokobylskij SA (1998) Tribe Alysiini. In: Ler PA (Ed.) Key to the Insects of Russian Far East. Dal'nauka, Vladivostok, 163–298. [In Russian]
- Belokobylskij SA (2005) Eastern Palaearctic species of the parasitic wasps of the genus *Aspilota* Foerster (Hymenoptera, Braconidae, Alysiinae). Species with developed mesoscutal pit. Entomologicheskoe Obozrenie 84(3): 610–641. [In Russian]
- Belokobylskij SA, Tobias VI (2007) Alysiinae. In: Lelej AS (Ed.) 'Key to the insects of Russia Far East. Vol. IV. Neuropteroidea, Mecoptera, Hymenoptera. Pt 5.' Dalnauka, Vladivostok, 9–133. [In Russian]
- Belokobylskij SA, Kostromina TS (2011) Two late-spring braconid genera of the family Alysiinae (Hymenoptera: Braconidae) new for the fauna of Russia. Zoosystematica Rossica 20(1): 85–95.
- Cameron P (1900) Descriptions of new genera and species of Hymenoptera. Annals and Magazine of Natural History 7(6): 530–541. https://doi.org/10.1080/00222930008678422
- Cameron P (1905) Description of a new genus and species of Braconidae from Cape Colony. Entomologist 38: 268–269.
- Cameron P (1910) On some Asiatic species of the Braconid subfamilies Rhogadinae, Agathinae, and Macrocentrinae and of the Alysiidae. Wiener Entomologische Zeitschrift 29: 1–10.
- Cameron P (1911) On the parasitic Hymenoptera collected by Mr. A.J.T. Janse, Transvaal. Annals of the Transvaal Museum 2: 173–217.
- Chen JH, Wu ZS (1994) The Alysiini of China: (Hymenoptera: Braconidae: Alysiinae). China Agricultural Press, Fuzhou, 1–218. [In Chinese, with summary in English]
- Chou LY (1981) A preliminary list of Braconidae (Hymenoptera) of Taiwan. Journal of Agricultural Research. China 30(1): 71–88.
- Curtis J (1826) British Entomology; being illustrations and descriptions of the genera of insects found in Great Britain and Ireland 3: 120, 141.
- Guerrieri E, Giorgini M, Cascone P, Carpenito S, van Achterberg C (2016) Species diversity in the parasitoid genus *Asobara* (Hymenoptera: Braconidae) from the native area of the fruit fly pest *Drosophila suzukii* (Diptera: Drosophilidae). Plos ONE 11(2): e0147382. https://doi.org/10.1371/journal.pone.0147382
- Fahringer J (1929) In: Seitner M. "Chortophila laricicola Karl, die Lärchen-zapfen und Samenfliege, und ihre Feinde: Parasiten und Räuber." Zentralblatt für das Gesamte Forstwesen 55: 153–167.

- Fischer M (1962) Das Genus *Synaldis* Foerster. Mitteilungen aus dem Zoologischen Museum in Berlin 38: 1–21.
- Fischer M (1966a) Über gezüchtete Braconiden aus Europa (Hymenoptera). Zeitschrift für Angewandte Entomologie 58: 323–339. https://doi.org/10.1111/j.1439-0418.1966. tb04349.x
- Fischer M (1966b) Studien über Alysiinae (Hymenoptera, Braconidae). Annalen des Naturhistorischen Museums in Wien 69: 177–205.
- Fischer M (1967) Seltene Alysiinae aus verschiedenen Erdteilen. Annalen des Naturhistorischen Museums in Wien 70: 109–138.
- Fischer M (1970a) Die Alysiini der Steiermark (Hymenoptera, Braconidae). Mitteilungen der Abteilung für Zoologie am Landesmuseum Joanneum 34: 1–44.
- Fischer M (1970b) Zur Kenntnis der europäischen *Phaenocarpa*-Arten mit besonderer Berücksichtigung der Fauna Niederösterreichs (Hymenoptera, Braconidae, Alysiinae). Zeitschrift für Angewandte Zoologie 57: 409–498.
- Fischer M (1973a) Redeskriptionen von Alysiinen (Hymenoptera, Braconidae). Annalen des Naturhistorischen Museums in Wien 77: 245–261.
- Fischer M (1973b) Einige Proben aus den Ötztaler Alpen als Beispiel für die Formenvielfalt bei der Gattung *Aspilota* Foerster (Hym., Braconidae, Alysiinae). Berichte des Naturwissenschaftlich-Medizinischen Vereins in Innsbruck 60: 95–129.
- Fischer M (1973c) Aspilota-wespen aus der weiteren Umgebung von Admont (Hym., Braconidae, Alysiinae). Mitteilungen der Abteilung für Zoologie am Landesmuseum Joanneum 2: 137–167.
- Fischer M (1975) Alysiinen-Wespen aus der Umgebung von Hüttenberg in Kärnten (Hymenoptera, Braconidae, Alysiinae). Carinthia 2: 303–342.
- Fischer M (1976a) Über zwei exotische Alysiinen-Wespen aus dem *Aspilota*-Komplex (Hymenoptera, Braconidae, Alysiinae). Zeitschrift der Arbeitsgemeinschaft Österreichischer Entomologen 27(3/4): 115–119.
- Fischer M (1976b) Erste Nachweise von *Aspilota*-Wespen in Burgenland (Hymenoptera, Braconidae, Alysiinae). Annalen des Naturhistorischen Museums in Wien 80: 343–410.
- Fischer M (1993) Zur Formenvielfalt der Kieferwespen der Alten Welt: Über die Gattungen *Synaldis* Foerster, *Trisynaldis* Fischer und *Kritscherysia* Fischer gen. n. (Hymenoptera, Braconidae, Alysiinae). Annalen des Naturhistorischen Museums in Wien 94/95B: 451–490.
- Fischer M (1995) Über die altweltlichen *Orthostigma*-Arten und Ergänzungen zur *Aspilota-*Gattungsgruppe (Hymenoptera, Braconidae, Alysiinae). Linzer Biologische Beiträge 27(2): 669–752.
- Fischer M (2003) Ein Beitrag zur Kenntnis der Gattungen *Synaldis* Foerster und *Adelphenaldis* Fischer, Gen. Nov. (Hymenoptera, Braconidae, Alysiinae). Linzer Biologische Beiträge 35(1): 19–74.
- Fischer M (2006) Neue Kieferwespen aus der Sammlung des Biologiezentrums des Oberösterreichischen Landesmuseums in Linz und Mitteilungen über andere Arten (Hymenoptera, Braconidae, Alysiinae). Linzer Biologische Beiträge 38/1: 605–651.
- Foerster A (1863) Synopsis der Familien und Gattungen der Braconiden. Verhandlungen des Naturhistorischen Vereins der Preussischen Rheinlande und Westfalens 19: 225–288.
- Gahan AB (1925) A second lot of parasitic Hymenoptera from the Philippines. Philippine Journal of Science 27: 83–109.

- Haliday AH (1833) An essay on the classification of the parasitic Hymenoptera of Britain, which correspond with Ichneumones minuti of Linnaeus. Entomological Magazine 1(iii): 259–276, 333–350.
- Haliday AH (1838) Essay on parasitic Hymenoptera. Entomological Magazine 5(3): 209–249.
- He JH, Chen XX, Fan JJ, Li Q, Liu CM, Lou XM, Ma Y, Wang SF, Wu YR, Xu ZH, Xu WA, Yao J (2004) Hymenopteran insect fauna of Zhejiang. Science Press, Beijing 1373: 354–816. [In Chinese with English summary]
- Hedqvist KJ (1973) Two new species of Alysiinae Först. from North Sweden (Hym., Ichneumonoidea, Braconidae). Entomologisk Tidskrift 94: 91–93.
- Königsmann E (1972) Zur Kenntnis verschiedener Gattungen der Alysiinae nebst Beschreibung der neuen Gattung *Paraorthostigma* (Hymenoptera, Braconidae). Deutsche Entomologische Zeitschrift 19(I/III): 21–30.
- Königsmann E (1960) Revision der paläarktischen Arten der Gattung *Idiasta*. 3. Beitrag zur systematischen Bearbeitung der Alysiinae (Hymenoptera: Braconidae). Beiträge zur Entomologie 10(5/6): 624–654.
- Königsmann E (1959) Revision der paläarktischen Arten der Gattung *Dapsilarthra*. 1. Beitrag zur systematischen Bearbeitung der Alysiinae (Hymenoptera: Braconidae). Beiträge zur Entomologie 9: 580–608.
- Königsmann E (1969) Beitrag zur Revision der Gattung *Orthostigma* (Hymenoptera, Braconidae). Deutsche Entomologische Zeitschrift 16 (I/III): 1–53.
- Marshall TA (1896) Les Braconides. In: André E (Ed.) Species des Hyménoptères d'Europe et d'Algérie 5 (1891): 1–635.
- Marsh PM (1979) Braconidae. Aphidiidae. Hybrizontidae. In: Krombein KV, Hurd Jr. PD, Smith DR, Burks BD (Eds) Catalog of Hymenoptera in America north of Mexico. Smithsonian Institution Press, Washington, 144–313.
- Nees von Esenbeck CG (1812) Icheneumonides Adesciti, in Genera et Familias Divisi. Magazin Gesellschaft Naturforschender Freunde zu Berlin 6(1812): 183–221.
- Nees von Esenbeck CG (1834) Hymenopterorum Ichneumonibus affinium monogarphiae, genera Europaea et species illustrantes. 1. Stuttgartiae et Tubingae 320.
- Nixon GEJ (1939) Notes on Alysiinae with descriptions of three new species (Hym., Braconidae). Proceedings of te Royal Entomological Society of London (B) 8(4): 61–67. https://doi.org/10.1111/j.1365-3113.1939.tb00493.x
- Papp J (1967) A synopsis of the *Phaenocarpa* Foerst. Species of the Oriental Region (Hymenoptera, Braconidae, Alysiinae). Reichenbachia 8: 139–157.
- Papp J (1968) A survey of the *Phaenocarpa* Förster species of the Carpathian Basin, Central Europe (Hymenoptera, Braconidae: Alysiinae). Beiträge zur Entomologie 18 (5/6): 569–603.
- Papp J (1969) *Conalysia* gen. n. and remarks on *Heratemis filosa* Walk. (Hym., Braconidae, Alysiinae). Opuscula Zoologica 9: 147–153.
- Papp J (1994) Braconidae (Hymenoptera) from Korea, XV. Acta Zoologica Academinae Scientiatum Hungaricae 40(2): 133–156.
- Panzer GWF (1799) Faunae Insectorum Germanicae. Heft 70–72.
- Peris-Felipo FJ, Jimenez-Peydro R (2011) Biodiversity within the subfamily Alysiinae (Hymenoptera, Braconidae) in the Natural Park Penas de Aya (Spain). Revista Brasileira de Entomologia 55(3): 406–410. https://doi.org/10.1590/S0085-56262011005000042

- Peris-Felipo FJ, Yari Z, van Achterberg C, Rakhshani E, Belokobylskij SA (2016) Review of species of the genus *Adelurola* Strand, 1928, with a key to species (Hymenoptera, Braconidae, Alysiinae). ZooKeys 566: 13–30. https://doi.org/10.3897/zookeys.566.6684
- Provancher L (1886) Additions et corrections au Volume II de la Faune Entomologique du Canada. Traitant des Hyménoptères. Québec, 475 pp.
- Ratzeburg JTC (1844) Die Ichneumonen der Forstinsecten in forstlicher und entomologischer Beziehung. Berlin, 224 pp.
- Rossi P (1807) Fauna Etrusca, sistens Insecta quae in Provinciis Florentina et Pisana. Tomus secundus. Helmstadii 577: 54–82.
- Ruthe JF (1854) Beiträge zur Kenntnis der Braconiden. Stettiner Entomologische Zeitung 15: 343–355.
- Sharma V (1978) Taxonomic studies on Indian Braconidae (Hymenoptera). Oriental Insects 12(1): 123–132. https://doi.org/10.1080/00305316.1978.10434561
- Shaw MR, Huddleston T (1991) Classification and biology of Braconid wasps (Hymenoptera: Braconidae). Handbooks for the identification of British Insects 7(11): 1–126.
- Stelfox AW (1941) Descriptions of five new species of Alysiidae (Hymenoptera) and notes on some others. Proceedings of the Royal Irish Academy 47(B): 1–16.
- Stelfox AW (1943) Description of *Aspilota dentifemur* sp. n. Proceedings of the Royal Irish Academy 49(B): 201–203.
- Stelfox AW (1944) *Phaenocarpa notabilis* sp.n. (Hym., Alysiidae) in Ireland. Entomologist's Monthly Magazine 80: 234–235.
- Strand E (1928) Miscellanea nomenclatorica zoologica et palaeontologica. I-II. Archiv für Naturgeschichte (A) 92(8) (1926): 30–75.
- Telenga NA (1935) Beiträge zur Kenntnis der Tribus Alysiini (Braconidae, Hymenoptera) aus USSR. Konowia 14: 186–190.
- Thomson CG (1895) LII. Bidrag till Braconidernas Kännedom. Opuscula Entomologica 20: 2141–2339.
- Tobias VI (1962) Contribution to the fauna of the subfamily Alysiinae (Hymenoptera, Braconidae) of the Leningrad region. Trudy Zoologicheskogo Insituta. Leningrad 31: 81–137. [In Russian]
- Tobias VI (1975) Two new species and a new genus of Braconids (Hymenoptera, Braconidae, Alysiinae) from Mongolia. Nasekomye Mongolii [Insects of Mongolia] 3: 306–309. [In Russian]
- Tobias VI, Jakimavicius A (1986) Alysiinae & Opiinae. In: Medvedev GS (Ed.) Opredelitel Nasekomych Evrospeiskoi Tsasti SSSR 3, Peredpontdatokrylye 4. Opr. Faune SSSR. Vol. 147, Section 3, Part 5 308: 7–231. [In Russian]
- Tobias VI (1990) Three new species of alysiine wasps (Hymenopera, Braconidae, Alysiinae) from Vietnam. Trudy Zoologicheskogo Instituta 209: 99–106. [In Russian]
- Tobias VI (2003) Species of the genus *Dinotrema* Foerster, 1862 (Hymenoptera, Braconidae, Alysiinae) without prescutellar pit and with smooth or only medially sculptured propodeum from Russia and adjacent territories. Entomologicheskoe Obozrenie 82(1): 138–156. [In Russian with English summary]
- van Achterberg C (1976) A new species of *Tanycarpa* Foerster from England (Hymenoptera, Braconidae, Alysiinae). Entomologische Berichten, Amsterdam 36: 12–15.
- van Achterberg C (1979) A revision of the subfamily Zelinae auct. (Hymenoptera, Braconidae). Tijdschrift voor Entomologie 122: 241–479.

- van Achterberg C, Bin F (1981) Notes on two species of *Dinotrema* Foerster (Hym., Braconidae, Alysiinae) with observations on the hymenopterous parasite-complex of *Spiniphora dorsalis* Becker (Dipt., Phoridae) in dead Helix spp. (Mollusca). Entomologische Berichten, Amsterdam 41: 104–112.
- van Achterberg C (1983a) Revisionary notes on the genera *Dapsilarthra* auct. and *Mesocrina* Foerster (Hymenoptera, Braconidae, Alysiinae). Tijdschrift voor Entomologie 126: 1–24.
- van Achterberg C, Ortega G (1983b) A new species of *Orthostigma* Ratzeburg from Tenerife (Insecta: Hymenoptera, Braconidae). Vieraea 12(1982): 121–127.
- van Achterberg C (1988a) Revision of the subfamily Blacinae Foerster (Hymenoptera, Braconidae). Zoologische Verhandelingen, Leiden 249: 1–324.
- van Achterberg C (1988b) The genera of the *Aspilota*-group and some descriptions of fungicolous Alysiini from the Netherlands (Hymenoptera: Braconidae: Alysiinae). Zoologische Verhandelingen, Leiden 247: 1–88.
- van Achterberg C (1998) *Bobekoides* gen. n. (Hymenoptera: Braconidae: Alysiinae) from South Africa. Zoologische Mededelingen, Leiden 72(9): 105–111.
- van Achterberg C (1993) Illustrated key to the subfamilies of the Braconidae (Hymenoptera: Ichneumonoidea). Zoologische Verhandelingen, Leiden 283: 1–189.
- van Achterberg C, Teiceira T, Oliveira L (2012) *Aphaereta ceratitivora* sp. n. (Hymenoptera, Braconidae), a new parastoid of *Ceratitis capitata* (Wiedemann) (Diptera, Tephritidae) from the Azores. ZooKeys 222: 1–9. https://doi.org/10.3897/zookeys.222.3618
- Walker F (1860) Characters of some apparently undescribed Ceylon insects. Annals and Magazine of Natural History (3) 5: 304–311.
- Wharton RA (1980) Review of the Nearctic Alysiini (Hymenoptera, Braconidae) with discussion of generic relationships within the tribe. University of California Press, Berkeley, U.S.A. 88: 1–112.
- Wharton RA (1984) Biology of the Alysiini (Hymenoptera: Braconidae), parasitoids of cyclor-rhaphous Diptera. Texas Agricultural Experimental Station. Technical Monograph 11: 1–39.
- Wharton RA, Chou LY (1985) Revision of the Taiwanese species of *Alloea* Haliday (Hymenoptera: Braconidae, Alysiinae). Journal of Agricultural Research of China 34(3): 352–367.
- Wharton RA (1986) The braconid genus *Alysia* (Hymenoptera): a description of the subgenera and a revision of the subgenus *Alysia*. Systematic Entomology 11: 453–504. https://doi.org/10.1111/j.1365-3113.1986.tb00538.x
- Wharton RA (1988) The braconid genus *Alysia* (Hym.): a revision of the subgenus *Anarcha*. Contributions of the American Entomological Institute 25: 1–69.
- Wharton RA (2002) Revision of the Australian Alysiini (Hymenoptera: Braconidae). Invertebrate Systematics 16 (1): 7–105. https://doi.org/10.1071/IT01012
- Wu ZS, Chen JH (1995a) Discovery of *Cratospila* Foerster from China (Hymenoptera, Braconidae). Journal of Fujian Agricultural University 24(3): 310–311.
- Wu ZS, Chen JH, Huang JC (1995b) New records of the genus *Phaenocarpa* Foerster (Hymenoptera: Braconidae) from China. Entomotaxonomia 17(4): 262.
- Wu ZS, Chen JH (1996) A new species of genus *Heratemis* Walker from Fujian province (Hymenoptera: Braconidae). Entomologia Sinica 3(1): 29–32. https://doi.org/10.1111/j.1744-7917.1996.tb00399.x
- Wu ZS, Chen JH (1998a) A new species of genus *Eudinostigma* (Hymenoptera: Braconidae) from China. Entomologia Sinica 5(1): 29–31. https://doi.org/10.1111/j.1744-7917.1998. tb00292.x

- Wu ZS, Chen JH (1998b) [Two newly recorded species of Alysiines from China (Hymenoptera: Braconidae).] Acta Zootaxonomica Sinica 23(2): 157. [In Chinese]
- Yaakop S, van Achterberg C, Ghani IBA (2009) *Heratemis* Walker (Hymenoptera: Braconidae: Alysiinae: Alysiini): revision and reconstruction of the phylogeny combining molecular data and morphology. Tijdschrift voor Entomologie 152(1): 3–64. https://doi.org/10.1163/22119434-900000268
- Yao JL, Kula RR, Wharton RA, Chen JH (2015a) Four new species of *Tanycarpa* (Hymenoptera, Braconidae, Alysiinae) from the Palaearctic Region and new records of species from China. Zootaxa 3957(2): 169–187. https://doi.org/10.11646/zootaxa.3957.2.2
- Yao JL, Kula RR, Wharton RA, Chen JH (2015b) A new species in the newly recorded genus *Trachyusa* (Hymenoptera: Braconidae: Alysiinae) from China. Zootaxa 3931(4): 579–584. https://doi.org/10.11646/zootaxa.3931.4.7
- Yu DS, van Achterberg C, Horstmann K (2016) Taxapad 2016, Ichneumonoidea 2015. Database on flash-drive. www.taxapad.com, Nepean, Ontario, Canada.
- Zheng ML, Chen JH, Yang JQ (2012) A new species in the genus *Hylcalosia* (Hymenoptera: Braconidae) from China. Entomotaxonomia 34(2): 453–458.
- Zhu JC, van Achterberg C, Chen XX (2017) A new genus and subgenus of Alysiini from China, with the description of two new species (Hymenoptera, Braconidae, Alysiinae). Zootaxa 4272(3): 360–370. https://doi.org/10.11646/zootaxa.4272.3.2
- Zaykov AN, Fischer M (1982) On three extraordinary Alysiinae from Bulgaria (Hym., Braconidae). Acta Zoologica Bulgarica 19: 70–74.
- Zetterstedt JW (1838) Insecta Lapponica: 358–408. Sectio secunda. Hymenoptera. Lipsiae.